









**Procedure**

1. Vortex the 500 µl reagent for 1 min to obtain a homogeneous mixture.
2. Add 0.5 µl of the reagent to each well of a 96-well plate (final volume 100 µl).
3. Place the microplate on a magnetic rack and incubate for 5 min at room temperature (20-25 °C).
4. Remove the liquid by pipetting it into a waste container. Discard the liquid.
5. Immediately add 100 µl of the sample to each well.
6. Place for 30 seconds.
7. Remove the liquid by pipetting it into a waste container. Discard the liquid.
8. Place for 20 seconds.
9. Remove the liquid by pipetting it into a waste container. Discard the liquid.
10. Place for 20 seconds.
11. Remove the liquid by pipetting it into a waste container. Discard the liquid.
12. Place for 20 seconds.
13. Remove the liquid by pipetting it into a waste container. Discard the liquid.
14. Place for 20 seconds.
15. Remove the liquid by pipetting it into a waste container. Discard the liquid.

**Fluorometric measurement of DNA concentration**

**Background**

Accurate measurement of double-stranded DNA concentration is essential for many molecular biology applications. The most common method for measuring DNA concentration is by using a spectrophotometer. However, this method is not always accurate, especially for low concentrations of DNA. Fluorometric measurement of DNA concentration is a more accurate method, especially for low concentrations of DNA. This method involves the use of a fluorescent dye that binds to DNA and emits light when excited. The intensity of the emitted light is proportional to the concentration of DNA.

**Equipment**

- OxiA 4 Fluorometer (meridian, Q3228)
- Microplates

Dear Gratiela Gradisteanu,

## **Wellcome Connecting Science: CRISPR Screening and Analysis**

**21/09/2025 – 26/09/2025**

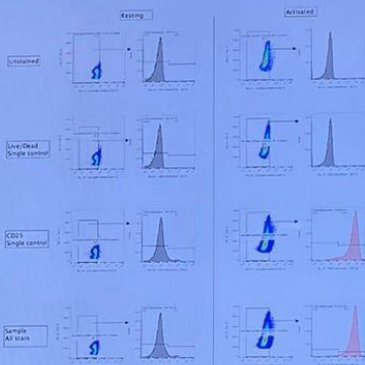
Congratulations! I am pleased to confirm that you have been selected to attend this event. The bioinformatics portion of the course will be held virtually on Monday 28 July - Friday 1 August 2025. Please be aware all times are British Summer Time, you should take this into account when planning your attendance if you are based in a different time zone. The laboratory course will be held onsite at the Wellcome Genome Campus from Sunday 21 - Friday 26 September 2025.

### **Acceptance**

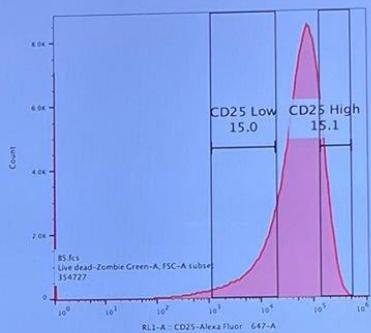
I am pleased to confirm you have been awarded a bursary towards your fees. This has been reflected in the invoice. Please note this is not transferable and we cannot offer any funding to cover travel expenses or other related costs.

## Demonstration

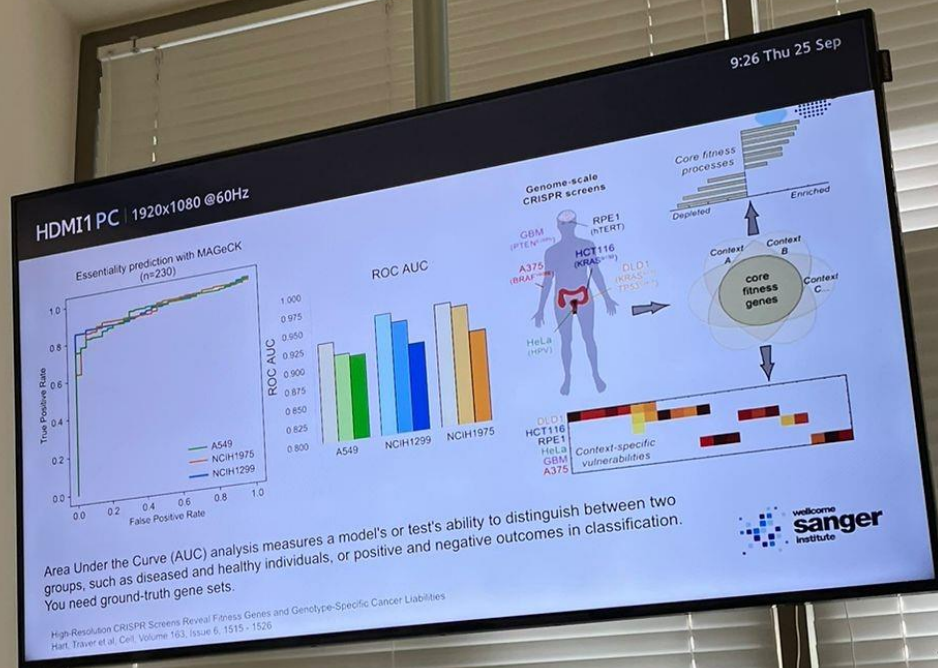
### Marker expression



### Sorting strategy



Open Targets



## Quality control



1

### Measure growth

Monitor OD600, growth rate, and biomass yield  
→ avoid severe fitness penalty



2

### Quantity SCFAs

GC-MS or HPLC to measure acetate, butyrate, propionate, lactate, ethanol.  
Expect butyrate ↑ 2-5x, ethanol ↓ 30-50%



3

### Redox balance

If NADH accumulates (detected by redox dyes, NADH/NAD<sup>+</sup> assays, or stunted growth), then Reduce CRISPRa strength on *bcd/etfAB\_R*



4

### Stability tests

Passage engineered strain over ≥10 generations  
ensure transgene persistence